

THE EFFECT OF CANCER-ASSOCIATED FIBROBLASTS ON ORAL SQUAMOUS CELL CARCINOMA

**Sadaf Rafiq¹, Sayed Hassan Raza Shah², Muhammad Ayub³, Salman
Khursheed⁴, Mohammad Usman Jan^{5a}, Kainat^{5b}, Kinza Taqdees⁶, Shan
Muzamail⁷**

¹Institute of Biological Sciences, Gomal University, D.I.K, Pakistan

Email: Sadaf.dcma.d.i.khan@gmail.com

²Eugene Applebaum College of Pharmacy and Health Sciences, Wayne State University, Detroit, Michigan 48202, Email: hk4165@wayne.edu

³ Department of Biological Sciences, The Superior University, Lahore, Pakistan, Email: ayub1141999@gmail.com

⁴Khyber College of Dentistry, Peshawar, Pakistan,
Email: salmankhursheed0011@gmail.com

^{5a}Department of Zoology, Abdul Wali Khan University, Mardan

Email: usmanjan@awkum.edu.pk

^{5b}Department of Zoology, Abdul Wali Khan University, Mardan

Email: kainatmeer3443@gmail.com

⁶ King Edward Medical University, Pakistan. Email: kinzataqdees77@gmail.com

⁷Graduate Institute of Nutrition, China University, Taichung, Taiwan

Email: shanmuzamail23@gmail.com

Corresponding Authors: Sadaf Rafiq, Institute of Biological Sciences,
Gomal University, D.I.K, Pakistan,

Email: Sadaf.dcma.d.i.khan@gmail.com

Shan Muzamail, Graduate Institute of Nutrition, China University, Taichung,
Taiwan, Email: shanmuzamail23@gmail.com

ABSTRACT

Cancer-associated fibroblasts (CAFs) are pivotal components of the tumor microenvironment (TME) that critically influence the progression and therapeutic resistance of oral squamous cell carcinoma (OSCC). This review provides an in-depth synthesis of current knowledge regarding the diverse roles of CAFs in OSCC, emphasizing their contributions to tumor growth, angiogenesis, metastasis, immune modulation, and drug resistance. CAFs, which originate from normal fibroblasts through activation processes such as TGF- β signaling and epithelial-mesenchymal transition, exhibit distinct markers including α -smooth muscle actin (α -SMA), fibroblast activating protein (FAP), and fibroblast specific protein-1 (FSP-1). Unlike their normal counterparts, CAFs secrete a complex array of cytokines and growth factors, such as IL-6, IL-8, vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), and hepatocyte growth factor (HGF), which activate oncogenic pathways including EGFR, Wnt/ β -catenin, Hippo, and JAK2-STAT3. These signaling networks facilitate not only enhanced tumor cell proliferation and invasion, but also the induction of epithelial-mesenchymal transition, thereby promoting metastasis. In addition, CAF-derived factors stimulate angiogenesis by recruiting endothelial progenitor cells and remodeling the extracellular matrix to support neovascularization. CAFs further contribute to an immunosuppressive TME by inducing T cell apoptosis and promoting M2 macrophage polarization, which impairs antitumor immunity. Notably, CAFs are implicated in resistance to conventional chemotherapies, underscoring their role in treatment failure. Targeting CAFs or their downstream effectors represents a promising therapeutic strategy to overcome drug resistance and improve patient outcomes. Overall, this review highlights the multifaceted impact of CAFs in OSCC and advocates for the development of CAF-directed therapies as an integral component of comprehensive cancer management. Future studies should aim

to elucidate the molecular heterogeneity of CAFs and to develop innovative strategies that effectively target their tumor-promoting functions without compromising normal tissue integrity.

KEYWORDS: Cancer-associated fibroblasts (CAFs); Oral squamous cell carcinoma (OSCC); Tumor microenvironment (TME); Epithelial-mesenchymal transition (EMT)

INTRODUCTION:

The incidence rate of cancer has been increasing rapidly ^[1]. Although significant progress has been made in studying the mechanisms underlying the occurrence and development of different types of cancer, addressing cancer related issues through these mechanisms still poses certain challenges ^[2]. The recurrence, metastasis, and tumor microenvironment (TME) of tumors remain three key and unresolved issues that hinder clinical treatment of cancer ^[3, 4]. The components of TME, such as cancer associated fibroblasts (CAFs), are activated fibroblasts in the tumor matrix and are associated with the occurrence and progression of malignant tumors ^[5]. Related studies have found that CAFs contribute to tumor growth and proliferation, angiogenesis, invasion and metastasis, and resistance to treatment ^[6]. Because CAFs are one of the most abundant matrix components in TME, they can serve as important therapeutic targets in many solid tumors ^[4]. Some of the solid tumors include head and neck cancer ^[6-8], breast cancer ^[9-11], lung cancer ^[12-15], Gastrointestinal, biliary cancers ^[16-18] and Urogenital system cancer ^[19, 20], etc. Our focus here is on head and neck squamous cell carcinoma, which is the seventh most common cancer worldwide and causes patient death ^[21, 22]. Therefore, it is important to understand and study how the TME of HNSCC promotes tumor progression ^[23, 24]. In this review, We have outlined the key role played by CAFs in head and neck squamous cell carcinoma, as well as some mechanisms of action, and may become potential targets for new treatment methods.

Tumor microenvironment:

The tumor microenvironment (TME) is the cellular environment in which tumors exist. The process of tumor progression and metastasis is the result of the joint action of non tumor cells and tumor cells ^[25]. In addition to tumor cells, TME mainly includes the following parts: (1) immune microenvironment (TIME): composed of various innate immune cells, such as tumor associated macrophages (TAMs), natural killer cells (NK) cells, neutrophils, mast cells, dendritic cells (DC), bone marrow-derived suppressor cells (MDSC), and adaptive immune cells, including CD4+T helper lymphocytes (Th), CD8+cytotoxic T cells NK-T cells $\gamma\delta$ T cell, regulatory T cell (Treg), and B cell composition^[26-28]. (2) Vascular components: The generation of tumor vascular networks is mainly achieved through the formation of new blood vessels, co selection and modification of existing blood vessels in tissues, and differentiation of bone marrow endothelium, all of which lead to heterogeneity of blood vessels within tumors^[25]. (3) Extracellular matrix (ECM): mainly composed of various collagen molecules, glycoproteins, and proteoglycans ^[29, 30]. (4) Non immune cells: such as cancer associated fibroblasts (CAFs) and mesenchymal stromal cells (MSCs). Among them, CAFs cells are the focus of this article's description.

Table 1: Components of the tumor microenvironment (TME)

Category	Components	Description
Immune Microenvironment (TIME)	- Tumor-associated macrophages (TAMs)	Various innate and adaptive immune cells interact with tumor cells.
	- Natural killer (NK) cells	

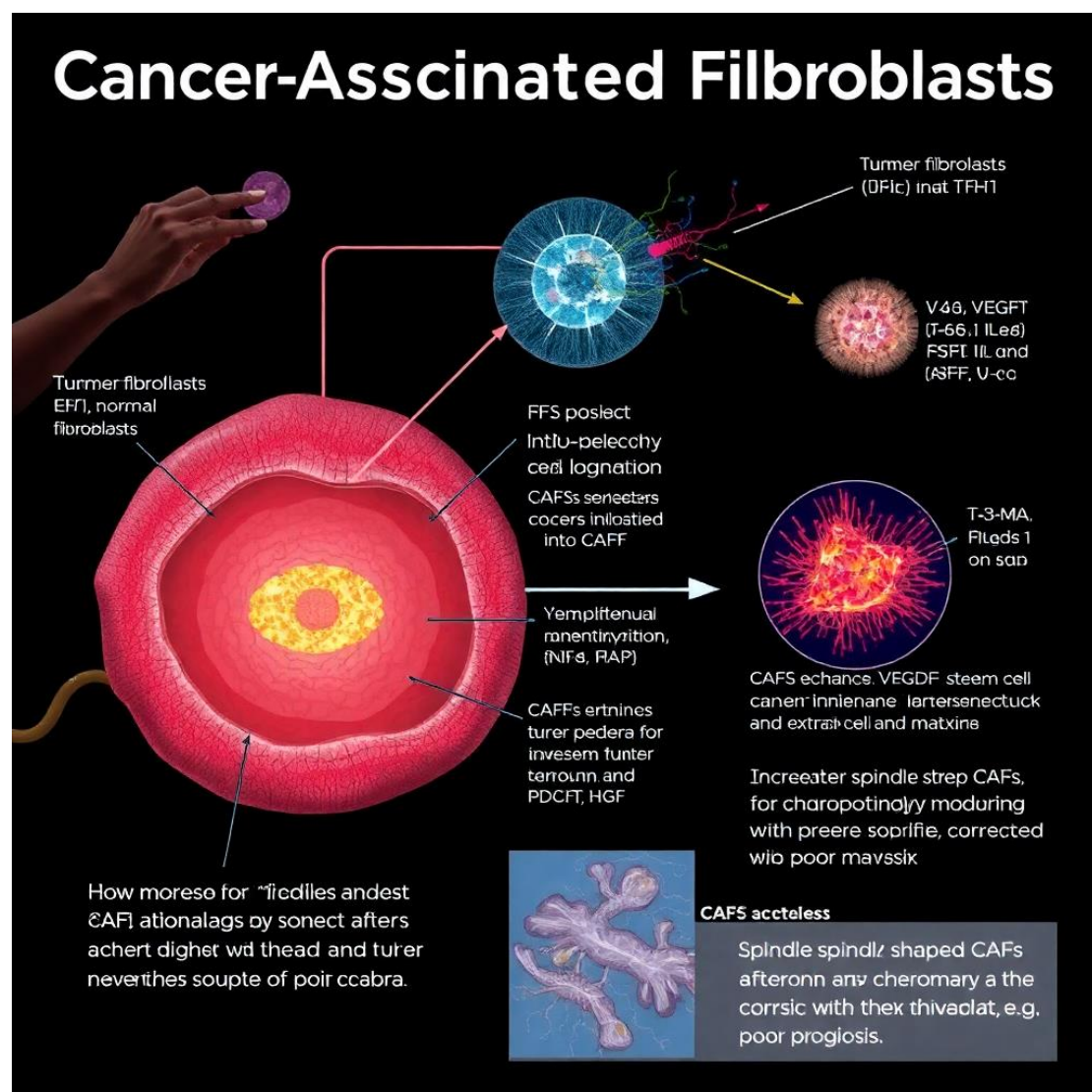
	- Neutrophils, mast cells, dendritic cells (DC)	
	- Bone marrow-derived suppressor cells (MDSCs)	
	- CD4+ T helper lymphocytes (Th)	
	- CD8+ cytotoxic T cells	
	- NK-T cells, $\gamma\delta$ T cells, regulatory T cells (Treg)	
	- B cells	
Vascular Components	- Tumor vascular networks	Formed through new blood vessel formation, co-selection and modification of existing vessels, and differentiation of bone marrow endothelium.
Extracellular Matrix (ECM)	- Collagen molecules	Provides structural and biochemical support.
	- Glycoproteins, proteoglycans	
Non-Immune Cells	- Cancer-associated fibroblasts (CAFs)	Play a significant role in tumor progression.
	- Mesenchymal stromal cells (MSCs)	

E-cadherin, resulting in the formation of mesenchymal morphology^[32, 33], Although normal fibroblasts (NFs) typically inhibit tumor formation^[34], cancer associated fibroblasts (CAFs) can significantly promote tumor development^[35]. Compared with NFs, CAFs not only increase cell proliferation, but also promote the production of extracellular matrix and the secretion of some special cytokines (such as stromal cell derived factor 1-SDF1, vascular endothelial growth factor VEGF, platelet derived growth factor PDGF, and hepatocyte growth factor HGF)^[36]. Transforming growth factor-1 (TGF) secreted by stromal cells and tumor cells- β 1) Is the main factor promoting the mobilization and activation of NFs into CAFs^[37, 38]. TGF- β 1. Activate fibroblasts to transform into CAFs and express smooth muscle actin through SMAD dependent or non dependent pathways (α - SMA, Periosteal Protein (POSTN) α - Fibroblast activating protein (α - FAP) and fibroblast specific protein-1 (FSP-1), and produce type I collagen^[39]. However, CAFs can not only express α - SMA α FAP and FSP-1, which can also produce vascular endothelial growth factor (VEGF) and cytokines such as IL-6 and IL-8^[40]. Research has shown that in TGF- β Under the influence of, cancer cells, especially cancer stem cells, can be a source of CAFs^[41]. The maintenance of stem cell characteristics is an important influencing factor for tumor formation; And the matrix can support and regulate the differentiation and proliferation of tumor stem cells^[42]. Fibroblasts are the main component of TME and regulate the occurrence and development of tumors by releasing various cytokines and interacting with various cells such as tumor cells^[4, 43].

In human head and neck squamous cell carcinoma samples, CAFs typically express myofibroblast markers α - SMA, showing spindle like cells^[44-46], Rarely co-expressed with other biomarkers^[47, 48]. After chemotherapy α - The expression of SMA may also increase^[44]. Studies have shown that the higher the density of CAFs, the worse their clinical manifestations (such as TNM staging, cervical lymph node metastasis, vascular

infiltration, postoperative recurrence, etc.)^[46, 49-51], the worse the prognosis after treatment. ^[46, 49, 52, 53]

TGF- β It has been proven to increase the cell strength of NFs and CAFs; It also increases the ability of CAFs to elongate and diffuse, which has never occurred in fibroblasts without activation; It also enhances the invasiveness of the tumor matrix ^[54].



The effect of CAFs on HNSCC.

Promoting tumor growth:

Unlike normal fibroblasts, CAFs can promote the growth and malignancy of non tumor epithelial cells Sexual transformation; This function was first discovered in a human prostate cancer mouse model ^[55]. It is widely believed that cancer cells and stromal cells (such as CAFs) dynamically co evolve during tumor progression ^[42]. In a three-dimensional co culture model, normal breast fibroblasts inhibit the growth of breast tumors and the transformation of breast epithelial cells, while induction by CAF promotes the transformation of normal breast epithelial cell lines (MCF10A and EIII8 cells, respectively)^[56].

Table 2: CAFs can lead to an increase in the malignancy of HNSCC, promoting tumor progression and metastasis, as shown in the table

Related signaling pathways	Impact on tumors	drug
EGFR	EGFR gene is amplified, overexpressed, or mutated in HNSCCs ^[57, 58] . The collective invasion of squamous cell carcinoma cells (SCCs) is driven by EGF signals ^[59]	Verapamil, Diltiazem ^[59]
WNT	Cyclins produced and secreted in large quantities by CAFs in HNSCC, through typical Wnt/ β -The catenin signaling pathway promotes CSC phenotype and promotes tumor progression and metastasis in HNSCC ^[60]	Erotinib ^[60]

Hippo	Transcription factor YAP is Gemcitabine ^[63] activated in CAFs ^[61] , YAP activation in the matrix is further enhanced in the surrounding tumor areas of advanced cancers such as breast cancer and squamous cell carcinoma ^[62]
NOTCH3	The expression of NOTCH3 in Taristazumab ^[64] CAFs is significantly correlated with microvascular density in cancer stroma, and the expression of NOTCH3 in CAFs is associated with poor prognosis in OSCC patients ^[64]
JAK2-STAT3	The up-regulated protein XX recombines cancer related fibroblasts and promotes the invasion of oral squamous cell carcinoma through the JAK2-STAT3 pathway ^[65]
IL-1 β	IL-1 β Inducing the production of Elotinib, Gifitinib, and CXCL1, which in turn activates Afatinib ^[66] EGFR through CXCR2, leading to autocrine proliferation of oral malignant precursor cells ^[66]
Hedgehog	CAFs are not only potential sources XX of HH ligands in tumor stroma, but may also respond to HH signaling through nuclear GLI-1 activation ^[67]

lncRNAH19/miR-675-5p/PFKFB3 Reprogramming of glycolysis in Glycolytic enzyme cancer related fibroblasts promotes inhibitors ^[68] the growth of oral cancer through the lncRNA H19/miR-675-5p/PFKFB3 signaling pathway ^[68]

Later, two different subpopulations of CAF-N (normal) and CAF-D (different) were described in human oral squamous cell carcinoma (OSCC). CAF-N secretes hyaluronic acid (HA) and matrix metalloproteinases (MMPs), promotes tissue invasion of cancer cells and fibroblasts, and produces ECM rich in HA and immunosuppression, while CAF-D is TGF- β Source of induction of EMT in cancer cells and promotion of cell migration ^[69, 70]. CAFs induce an increase in vimentin and a decrease in e-cadherin expression in oral cancer cell lines, indicating the occurrence of EMT ^[47, 71, 72]. Epithelial regulatory proteins are a growth factor in the EGF family, and their overexpression can induce phosphorylation of JAK2/STAT3 and secretion of IL-6 in NFs, which plays an important role in promoting EMT in oral cancer cells ^[65]. CAFs activate EMT signals through various cytokines, promote tumor cell invasion and metastasis, regulate the biological behavior of tumor cells, and thus affect the growth of OSCC ^[73, 74].

Promoting angiogenesis:

If new blood vessels are not formed to provide oxygen and nutrition, tumors cannot sustain their own growth in the body ^[75]. The key steps in tumor angiogenesis include the aggregation of endothelial progenitor cells and the migration of vascular endothelial cells ^[76]. CAFs enhance tumor angiogenesis by inducing the mobilization and aggregation of endothelial progenitor cells ^[35, 77]. And CAF can secrete various vascular growth factors, such as VEGF, PDGF, CXCL-12, or HGF, promoting ECM remodeling, proliferation of ECs, and aggregation of ECs and surrounding cells into the tumor ^{[78-}

^{80]}.PDGF and VEGF also have an autocrine effect on CAFs, further stimulating the production of other angiogenic factors such as IL-6, IL-8, and placental growth factor (PGF)^[81, 82].Studies have shown that in animal experiments, injecting CAFs and tumor cells subcutaneously into mice increases the rate of peripheral angiogenesis ^[83].Tumor angiogenesis is essential for tumor growth, invasion, and metastasis. Currently, the gold standard for describing tumor angiogenesis is histological MVD technology ^[84].Wang et al. found the characteristics of CAFs in head and neck squamous cell carcinoma (HNSCCs) ^[76];They found that in the matrix of nasopharyngeal carcinoma (NPC), fibroblasts α - The expression of SMA is significantly higher; In NPC cells, the immune response intensity of SDF-1 and CXCR4 secreted by CAFs is also high, indicating the presence of endothelial progenitor cells in both cancer and stromal cells of NPC. Stromal cells enhance neovascularization in a VEGF - and SDF-1 dependent manner. Promoting tumor proliferation and metastasis:

CAF's secrete multiple factors, such as TGF- β 1. CXCL-12, FGF, POSTN, osteopontin (OPN), hepatocyte growth factor (HGF), IL-6, and IL-22 directly stimulate tumor proliferation and growth through their respective signaling pathways; For example: integer/FAK src (POSTN), Wnt/ β - Catenin (HGF and OPN), PI3K/mTOR (CXCL-12, HGF and IL-22), MAPK (IL-6, TGF- β And FGF) or Hippo (EVs)^[85-92].Ramos Vega et al. demonstrated that, α - There is a significant association between SMA positive CAF cells and advanced clinical stages and poorly differentiated tumors of laryngeal cancer and HNSCC cancer ^[5].The possible mechanisms by which CAFs promote transfer are also diverse; CAFs can secrete cytokines and chemokines to specifically support tumor progression; For example, the chemokines of CCL5 acting on cancer cells can promote invasion and metastasis ^[93].For advanced CAFs, also as a subtype of CAFs, it can secrete excessive IL-8, which is the mediator of the interaction between cancer cells and CAF, and promotes the invasion and metastasis of pancreatic cancer

cells ^[94].Late stage CAFs can also inhibit the immune response to tumors and reshape the extracellular matrix, promoting tumor invasion ^[95].

CAFs may also lead to bone invasion of head and neck tumors ^[71].Bone resorption usually involves the activation of osteoblasts through the interaction between the receptor activator of nuclear factor kappaB (RANK) and its osteoblast secretory ligand (RANKL); Bone protein growth hormone (OPG), as a receptor for RANKL, prevents excessive bone resorption by preventing RANK-RANKL interactions ^[96,97].Oral cancer cells and stromal cells both secrete RANKL, but CAFs have been shown to promote osteoclast resorption to a greater extent in vitro and induce multinucleation of mouse macrophages, which may be osteoblasts ^[96].

Research has shown that when cancer cells carry their own ectopic stromal cell fragments, their survival ability after entering the circulation is greater; In addition, when these circulating tumor cells with matrix components and cancer related fibroblasts are depleted, the number of lung metastases is significantly reduced and the survival rate is prolonged; This indicates the importance of CAFs in the development of transfer ^[94, 95].

Table 3: Role of cancer-associated fibroblasts (CAFs) in tumor angiogenesis, proliferation, metastasis, and bone invasion

Category	Mechanisms	Key Molecules/Factors	Effects	References
Promoting Angiogenesis	Aggregation of endothelial progenitor cells	CAFs mobilize endothelial progenitor cells	Enhances tumor angiogenesis	[35, 77]
	Migration of vascular	VEGF, PDGF, CXCL-12, HGF	Promotes ECM remodeling, endothelial cell	[78-80]

	endothelial cells		proliferation, and tumor vascularization	
	Autocrine signaling in CAFs	IL-6, IL-8, PGF	Further increases angiogenic factor production	[81, 82]
	Experimental validation	Injection of CAFs and tumor cells in mice	Increases peripheral angiogenesis	[83]
	Tumor angiogenesis assessment	Histological MVD technology	Gold standard for describing angiogenesis	[84]
	CAFs in head and neck cancers (HNSCC, NPC)	α -SMA, SDF-1, CXCR4, VEGF	Enhances neovascularization via VEGF and SDF-1	[76]
Promoting Tumor Proliferation and Metastasis	Secretion of growth factors	TGF- β 1, CXCL-12, FGF, POSTN, OPN, HGF, IL-6, IL-22	Stimulates tumor proliferation via multiple signaling pathways	[85-92]
	Key signaling pathways	Integrin/FAK/Src, Wnt/ β -catenin, PI3K/mTOR, MAPK, Hippo	Drives tumor proliferation and metastasis	[85-92]
	CAFs in laryngeal and	α -SMA positive CAF cells	Associated with advanced clinical	[5]

	HNSCC cancers		stages and poor differentiation	
	Secretion of cytokines and chemokines	CCL5, IL-8	Promotes invasion and metastasis, particularly in pancreatic cancer	[93, 94]
	Late-stage CAFs	IL-8, ECM remodeling	Inhibits immune response and promotes tumor invasion	[95]
Bone Invasion by Tumors	Bone resorption mechanism	RANK-RANKL interaction	Activates osteoclasts and promotes bone invasion	[96, 97]
	Regulation of bone degradation	OPG	Inhibits excessive bone resorption by blocking RANK-RANKL interaction	[96, 97]
	CAFs' role in bone invasion	RANKL secretion, osteoclast resorption induction	Promotes osteoclast activation and multinucleation of macrophages	[96]
Survival and Metastatic Potential	Circulating tumor cells (CTCs)	Tumor cells with CAF-associated stroma	Increased survival in circulation	[94, 95]
	Depletion of CAFs in CTCs	Reduced lung metastases, prolonged survival	Highlights importance of CAFs in metastasis	[94, 95]

The effect of CAFs on immune cells in tumor microenvironment:

In HNSCC, CAFs can directly affect ^[98]And indirect impacts ^[49]Immune cells. On the one hand, CAFs induce apoptosis of CD4 and CD8T cells, while increasing the proportion and migration of regulatory T cells (Tregs) that inhibit T cell anti-tumor response ^[98, 99]. Impaired T cell proliferation may be due to the secretion of immunosuppressive cytokines by CAFs, as well as the expression of B7 family, B7H1 (PDL1), and B7DC (PDL2) ^[98]. On the other hand, CAFs, whether acting alone or in collaboration with cancer cells, can cause monocytes to differentiate into the M2 macrophage phenotype of the original tumor ^[49, 71, 100]. Then, these macrophages may secrete TGF- β , IL-10 and arginase I have inhibitory effects on T cells ^[49]. Once macrophages differentiate into the M2 phenotype, they also enhance growth, invasion, migration, and CSC characteristics to affect oral cancer cells ^[97]. Similar effects of CAF have also been observed in esophageal squamous cell carcinoma, where the HGF released by CAF and its receptor MET interact with tumor cells to enhance their invasiveness ^[101]. Dendritic cells (DCs), as known cells with strong antigen-presenting function, are responsible for presenting tumor antigens and activating specific immune responses. CAFs secrete TGF- β And VEGF has the ability to inhibit antigen presentation to T cells, leading to an anti-tumor immune response ^[102].

There are two main roles of stromal cells (especially CAFs) in immune regulation of tumor tissue; On the first hand, CAFs promote tumor development by inducing a chronic inflammatory state in cancer cells; The second aspect is to promote the survival of tumors, which can alleviate the body's immune response to tumors, which is also a crucial aspect ^[42, 103, 104]. For HNSCC, in a study to investigate the relationship between CAFs and TAMs, Yu et al. demonstrated through immunohistochemical experiments that, α - The density of SMA positive and CAFs is closely related to CD-163 positive TAMs ^[105].

The impact of CAF on drug resistance:

Resistance to cancer treatment often leads to further exacerbation of tumors, and many studies have investigated the role of stromal cell CAFs in conferring treatment resistance based on their response to anti-tumor therapy ^[2]. Some research results in the literature have found a relationship between the resistance of stromal cells, especially CAFs, and various cancers, some of which include HNSCC's resistance to cetuximab ^[8]; Estrogen receptor (ER) negative breast cancer is resistant to 5-fluorouracil and cyclophosphamide (FEC) treatment ^[10]; Triple negative breast cancer is resistant to adriamycin ^[106]. Among them, in the study of HNSCC's resistance to cetuximab, Johansson et al. established a co culture model of HNSCC and CAFs, and the results showed that HNSCC and CAFs co culture were resistant to cetuximab treatment, and this effect was concentration dependent ^[8]. In breast, colorectal, and pancreatic tumors, CAFs can produce IL-6, IL-17A, PDGF, and insulin-like growth factor (IGF), which can activate NF of doxorubicin, 5-fluorouracil, and cisplatin- κ B and ERK pathways promote the stability of anti apoptotic proteins and the proliferation of cancer stem cells ^[29, 107-111]. Studies have shown that TGF secreted by CAFs- β Has a certain impact on tumors ^[112], So targeting TGF secreted by CAFs- β It can reduce the resistance of tumors. For HNSCC, survival of CAF can still be observed in the body after cisplatin and radiotherapy ^[113, 114]. And after treatment, it may induce the activation of advanced CAF phenotype in oral fibroblasts ^[115-117].

Treatment methods for CAFs:

The role of CAFs in promoting tumor occurrence, proliferation, and metastasis during the development of cancer makes them therapeutic targets for cancer intervention. However, the treatment of CAFs faces many obstacles and challenges. Due to the lack of specific CAF cell surface markers, this limits the direct action of drugs and makes it

difficult to accurately target CAFs without damaging normal tissues ^[2]. At present, there are two types of targeted treatments for CAFs, one is to directly act on CAFs (remove CAFs or inhibit their activity)^[118, 119] The second type indirectly affects CAFs (by inhibiting downstream effector molecules of CAFs and targeting extracellular matrix)^[120-122].

FAP is not only a surface marker activated by over 90% of human cancer fibroblasts, but also regulates the differentiation and proliferation of myofibroblasts ^[123]. FAP5-DM1 is a novel anti FAP monoclonal antibody (MAb) that can inhibit tumor growth for a long time in lung cancer and pancreas, and there are no toxic reactions observed even in head and neck cancer metastasis models ^[124]. FAP monoclonal antibody F19 has in vivo safety and has been used in phase I clinical trials in patients with rectal cancer and small cell lung cancer ^[125].

α -SMA, as a myofibroblast marker in CAFs, has been shown in an animal experiment to target α -SMA enhances the infiltration of immunosuppressive CD3⁺Foxp3⁺Treg cells into tumors, ultimately leading to increased tumor invasiveness and reduced animal survival rate ^[126].

Due to FAP and α -SMA is not entirely expressed by fibroblasts, which greatly hinders the accuracy of targeting CAFs using the above methods. Therefore, the potential source of cells targeting CAFs may be another method to reduce CAF in tumors; Endothelial cells are a potential source of CAFs, and phase III clinical trials are underway to target these CAF precursors with bevacizumab ^[127].

(CAFS) to congruently birorgete(CAF IMi) in tumor-association and metastases.



knowledge has laid a solid foundation for developing new treatment strategies for cancer treatment. In early studies, the difference between CAFs and normal fibroblasts could be distinguished by several typical markers, but the discovery of specific recognition markers for CAFs was not achieved. With continuous efforts and understanding of the molecular mechanisms underlying CAF pathology, many drugs targeting key regulatory factors are currently undergoing clinical and/or preclinical evaluations. With the development of single-cell sequencing and the advancement of cell specific new biomaterials and other technologies, we can selectively eliminate the ability to promote tumor CAF or reverse its tumor promoting activity, which may become an effective treatment method for cancer alone or in combination with other cancer treatment methods.

REFERENCES

- [1] TORRE L A, BRAY F, SIEGEL R L, et al. Global cancer statistics, 2012 [J]. CA Cancer J Clin, 2015, 65(2): 87-108.
- [2] CHEN X, SONG E. Turning foes to friends: targeting cancer-associated fibroblasts [J]. Nat Rev Drug Discov, 2019, 18(2): 99-115.
- [3] ZHOU B B, ZHANG H, DAMELIN M, et al. Tumour-initiating cells: challenges and opportunities for anticancer drug discovery [J]. Nat Rev Drug Discov, 2009, 8(10): 806-23.
- [4] KALLURI R. The biology and function of fibroblasts in cancer [J]. Nat Rev Cancer, 2016, 16(9): 582-98.
- [5] RAMOS-VEGA V, VENEGAS ROJAS B, DONOSO TORRES W. Immunohistochemical analysis of cancer-associated fibroblasts and podoplanin in head and neck cancer [J]. Med Oral Patol Oral Cir Bucal, 2020, 25(2): e268-e76.

- [6] RAUDENSKA M, SVOBODOVA M, GUMULEC J, et al. The Importance of Cancer-Associated Fibroblasts in the Pathogenesis of Head and Neck Cancers [J]. *Klin Onkol*, 2020, 33(1): 39-48.
- [7] ZHU Y, SHI C, ZENG L, et al. High COX-2 expression in cancer-associated fibroblasts contributes to poor survival and promotes migration and invasiveness in nasopharyngeal carcinoma [J]. *Mol Carcinog*, 2020, 59(3): 265-80.
- [8] JOHANSSON A C, ANSELL A, JERHAMMAR F, et al. Cancer-associated fibroblasts induce matrix metalloproteinase-mediated cetuximab resistance in head and neck squamous cell carcinoma cells [J]. *Mol Cancer Res*, 2012, 10(9): 1158-68.
- [9] MISHRA P J, MISHRA P J, HUMENIUK R, et al. Carcinoma-associated fibroblast-like differentiation of human mesenchymal stem cells [J]. *Cancer Res*, 2008, 68(11): 4331-9.
- [10] FARMER P, BONNEFOI H, ANDERLE P, et al. A stroma-related gene signature predicts resistance to neoadjuvant chemotherapy in breast cancer [J]. *Nat Med*, 2009, 15(1): 68-74.
- [11] WU H J, HAO M, YEO S K, et al. FAK signaling in cancer-associated fibroblasts promotes breast cancer cell migration and metastasis by exosomal miRNAs-mediated intercellular communication [J]. *Oncogene*, 2020, 39(12): 2539-49.
- [12] FENGZHOU LI S Z, YANWEI CUI, TAO GUO, JIAQI QIANG, QIANG XIE, WENDAN YU, WEI GUO, WUGUO DENG, CHUNDONG GU, TAIHUA WU α 1,6-Fucosyltransferase (FUT8) regulates the cancer-promoting capacity of cancer-associated fibroblasts (CAFs) by modifying EGFR core fucosylation (CF) in non-small cell lung cancer (NSCLC) [J]. *Am J Cancer Res*, 2020, 10(3): 816-37.
- [13] SANTOS A M, JUNG J, AZIZ N, et al. Targeting fibroblast activation protein inhibits tumor stromagenesis and growth in mice [J]. *J Clin Invest*, 2009, 119(12): 3613-25.

- [14]NATALIE C DIREKZE K H-D, ROSEMARY JEFFERY, TOBY HUNT, RICHARD POULSOM, DAHMANE OUKRIF, MALCOLM R ALISON, NICHOLAS A WRIGHT. Bone marrow contribution to tumor-associated myofibroblasts and fibroblasts [J]. CANCER RESERCH, 2004, 64(23): 8492-5.
- [15]HUANG C, XU J, LI Z. [Research Progress of Cancer-associated Fibroblasts in Lung Cancer] [J]. Zhongguo Fei Ai Za Zhi, 2020, 23(4): 267-73.
- [16]JAVIER VAQUERO , LYND A AOU DJEHANE , FOUASSIER L. Cancer-associated fibroblasts in cholangiocarcinoma [J]. Curr Opin Gastroenterol, 2020, 36(2): 63-9.
- [17]ZHANG H, DENG T, LIU R, et al. CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer [J]. Mol Cancer, 2020, 19(1): 43.
- [18]BAI Y P, SHANG K, CHEN H, et al. FGF-1/-3/FGFR4 signaling in cancer-associated fibroblasts promotes tumor progression in colon cancer through Erk and MMP-7 [J]. Cancer Sci, 2015, 106(10): 1278-87.
- [19]COLVIN E K, HOWELL V M, MOK S C, et al. Expression of long noncoding RNAs in cancer-associated fibroblasts linked to patient survival in ovarian cancer [J]. Cancer Sci, 2020, 111(5): 1805-17.
- [20]TENG F, TIAN W Y, WANG Y M, et al. Cancer-associated fibroblasts promote the progression of endometrial cancer via the SDF-1/CXCR4 axis [J]. J Hematol Oncol, 2016, 9(8).
- [21]CHOW L Q M. Head and Neck Cancer [J]. N Engl J Med, 2020, 382(1): 60-72.
- [22]ALSAHAFAI E, BEGG K, AMELIO I, et al. Clinical update on head and neck cancer: molecular biology and ongoing challenges [J]. Cell Death Dis, 2019, 10(8): 540.
- [23]PELTANOVA B, RAUDENSKA M, MASARIK M. Effect of tumor microenvironment on pathogenesis of the head and neck squamous cell carcinoma: a

systematic review [J]. *Mol Cancer*, 2019, 18(1): 63.

[24]SCHMITZ S, MACHIELS J P. Targeting the Tumor Environment in Squamous Cell Carcinoma of the Head and Neck [J]. *Curr Treat Options Oncol*, 2016, 17(7): 37.

[25]JUNTTILA M R, DE SAUVAGE F J. Influence of tumour micro-environment heterogeneity on therapeutic response [J]. *Nature*, 2013, 501(7467): 346-54.

[26]YANG F, WANG T, DU P, et al. M2 bone marrow-derived macrophage-derived exosomes shuffle microRNA-21 to accelerate immune escape of glioma by modulating PEG3 [J]. *Cancer Cell Int*, 2020, 20(93).

[27]AMY J PETTY Y Y. Tumor-associated macrophages: implications in cancer immunotherapy [J]. *Immunotherapy*, 2017, 9(3): 289-302.

[28]ZHANG D, ZHENG Y, LIN Z, et al. Equipping Natural Killer Cells with Specific Targeting and Checkpoint Blocking Aptamers for Enhanced Adoptive Immunotherapy in Solid Tumors [J]. *Angew Chem Int Ed Engl*, 2020, 59(29): 12022-8.

[29]LOUAULT K, LI R R, DECLERCK Y A. Cancer-Associated Fibroblasts: Understanding Their Heterogeneity [J]. *Cancers (Basel)*, 2020, 12(11):

[30]GIRARD C A, LECACHEUR M, BEN JOUIRA R, et al. A Feed-Forward Mechanosignaling Loop Confers Resistance to Therapies Targeting the MAPK Pathway in BRAF-Mutant Melanoma [J]. *Cancer Res*, 2020, 80(10): 1927-41.

[31]SUGIMOTO H, MUNDEL T M, KIERAN M W, et al. Identification of fibroblast heterogeneity in the tumor microenvironment [J]. *Cancer Biol Ther*, 2006, 5(12): 1640-6.

[32]FORINO M, TORREGROSSA R, CEOL M, et al. TGFbeta1 induces epithelial-mesenchymal transition, but not myofibroblast transdifferentiation of human kidney tubular epithelial cells in primary culture [J]. *Int J Exp Pathol*, 2006, 87(3): 197-208.

[33]OFT M, AKHURST R J, BALMAIN A. Metastasis is driven by sequential elevation of H-ras and Smad2 levels [J]. *Nat Cell Biol*, 2002, 4(7): 487-94.

- [34]G P DOTTO R A W, A ARIZA. Malignant transformation of mouse primary keratinocytes by Harvey sarcoma virus and its modulation by surrounding normal cells [J]. Proc Natl Acad Sci U S A, 1988, 85(17): 6389-93.
- [35]ORIMO A, GUPTA P B, SGROI D C, et al. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion [J]. Cell, 2005, 121(3): 335-48.
- [36]POLANSKA U M, ORIMO A. Carcinoma-associated fibroblasts: non-neoplastic tumour-promoting mesenchymal cells [J]. J Cell Physiol, 2013, 228(8): 1651-7.
- [37]ZHANG J, LIU J. Tumor stroma as targets for cancer therapy [J]. Pharmacol Ther, 2013, 137(2): 200-15.
- [38]L RønNOV-JESSEN O W P, V E KOTELIANSKY, M J BISSELL. The origin of the myofibroblasts in breast cancer. Recapitulation of tumor environment in culture unravels diversity and implicates converted fibroblasts and recruited smooth muscle cells [J]. J Clin Invest, 1995, 95(2): 859-73.
- [39]BHOWMICK N A, CHYTIL A, PLIETH D, et al. TGF-beta signaling in fibroblasts modulates the oncogenic potential of adjacent epithelia [J]. Science, 2004, 303(5659): 848-51.
- [40]EREZ N, TRUITT M, OLSON P, et al. Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting Inflammation in an NF-kappaB-Dependent Manner [J]. Cancer Cell, 2010, 17(2): 135-47.
- [41]NAIR N, CALLE A S, ZAHRA M H, et al. A cancer stem cell model as the point of origin of cancer-associated fibroblasts in tumor microenvironment [J]. Sci Rep, 2017, 7(1): 6838.
- [42]JOSHI R S, KANUGULA S S, SUDHIR S, et al. The Role of Cancer-Associated Fibroblasts in Tumor Progression [J]. Cancers (Basel), 2021, 13(6):
- [43]SAKAMOTO A, KUNOU S, SHIMADA K, et al. Pyruvate secreted from patient-

derived cancer-associated fibroblasts supports survival of primary lymphoma cells [J]. Cancer Sci, 2019, 110(1): 269-78.

[44] LIANG L, LUO H, HE Q, et al. Investigation of cancer-associated fibroblasts and p62 expression in oral cancer before and after chemotherapy [J]. J Craniomaxillofac Surg, 2018, 46(4): 605-10.

[45] VALACH J, FIK Z, STRNAD H, et al. Smooth muscle actin-expressing stromal fibroblasts in head and neck squamous cell carcinoma: increased expression of galectin-1 and induction of poor prognosis factors [J]. Int J Cancer, 2012, 131(11): 2499-508.

[46] VERED M, SHNAIDERMAN-SHAPIRO A, ZLOTOGORSKI-HURVITZ A, et al. Cancer-associated fibroblasts in the tumor microenvironment of tongue carcinoma is a heterogeneous cell population [J]. Acta Histochem, 2019, 121(8): 151446.

[47] LI H, ZHANG J, CHEN S W, et al. Cancer-associated fibroblasts provide a suitable microenvironment for tumor development and progression in oral tongue squamous cancer [J]. J Transl Med, 2015, 13(198).

[48] ZELTZ C, ALAM J, LIU H, et al. alpha11beta1 Integrin is Induced in a Subset of Cancer-Associated Fibroblasts in Desmoplastic Tumor Stroma and Mediates In Vitro Cell Migration [J]. Cancers (Basel), 2019, 11(6):

[49] HIDEYUKI TAKAHASHI K S, TAKESHI KUDO , MINORU TOYODA , KYOICHI KAIRA , TETSUNARI OYAMA , KAZUAKI CHIKAMATSU. Cancer-associated fibroblasts promote an immunosuppressive microenvironment through the induction and accumulation of protumoral macrophages [J]. Oncotarget, 2017, 8(5): 8633-47.

[50] BAGORDAKIS E, SAWAZAKI-CALONE I, MACEDO C C, et al. Secretome profiling of oral squamous cell carcinoma-associated fibroblasts reveals organization and disassembly of extracellular matrix and collagen metabolic process signatures [J]. Tumour Biol, 2016, 37(7): 9045-57.

- [51]SUN L P, XU K, CUI J, et al. Cancerassociated fibroblastderived exosomal miR3825p promotes the migration and invasion of oral squamous cell carcinoma [J]. Oncol Rep, 2019, 42(4): 1319-28.
- [52]PARAJULI H, TEH M T, ABRAHAMSEN S, et al. Integrin alpha11 is overexpressed by tumour stroma of head and neck squamous cell carcinoma and correlates positively with alpha smooth muscle actin expression [J]. J Oral Pathol Med, 2017, 46(4): 267-75.
- [53]MARSH D, SUCHAK K, MOUTASIM K A, et al. Stromal features are predictive of disease mortality in oral cancer patients [J]. J Pathol, 2011, 223(4): 470-81.
- [54]STYLIANOU A, GKRETSI V, STYLIANOPOULOS T. Transforming growth factor-beta modulates pancreatic cancer associated fibroblasts cell shape, stiffness and invasion [J]. Biochim Biophys Acta Gen Subj, 2018, 1862(7): 1537-46.
- [55]A F OLUMI G D G, S W HAYWARD, P R CARROLL, T D TLSTY, G R CUNHA. Carcinoma-associated fibroblasts direct tumor progression of initiated human prostatic epithelium [J]. Cancer Res, 1999, 59(19): 5002-11.
- [56]MALATHY P. V. SHEKHAR J W, STEVE J. SANTNER, ROBERT J. PAULEY, AND LARRY TAIT. Breast Stroma Plays a Dominant Regulatory Role in Breast Epithelial Growth and Differentiation: Implications for Tumor Development and Progression [J]. CANCER RESERCH, 2001, 61(4): 1320-6.
- [57]GULLICK W J. Prevalence of aberrant expression of the epidermal growth factor receptor in human cancers [J]. Br Med Bull, 1991, 47(1): 87-98.
- [58]ERJALA K, SUNDVALL M, JUNTILA T T, et al. Signaling via ErbB2 and ErbB3 associates with resistance and epidermal growth factor receptor (EGFR) amplification with sensitivity to EGFR inhibitor gefitinib in head and neck squamous cell carcinoma cells [J]. Clin Cancer Res, 2006, 12(13): 4103-11.
- [59]GRASSET E M, BERTERO T, BOZEC A, et al. Matrix Stiffening and EGFR

Cooperate to Promote the Collective Invasion of Cancer Cells [J]. *Cancer Res*, 2018, 78(18): 5229-42.

[60] YU B, WU K, WANG X, et al. Periostin secreted by cancer-associated fibroblasts promotes cancer stemness in head and neck cancer by activating protein tyrosine kinase 7 [J]. *Cell Death & Disease*, 2018, 9(11):

[61] FOSTER C T, GUALDRINI F, TREISMAN R. Mutual dependence of the MRTF-SRF and YAP-TEAD pathways in cancer-associated fibroblasts is indirect and mediated by cytoskeletal dynamics [J]. *Genes Dev*, 2017, 31(23-24): 2361-75.

[62] CALVO F, EGE N, GRANDE-GARCIA A, et al. Mechanotransduction and YAP-dependent matrix remodelling is required for the generation and maintenance of cancer-associated fibroblasts [J]. *Nat Cell Biol*, 2013, 15(6): 637-46.

[63] PROVENZANO P P, CUEVAS C, CHANG A E, et al. Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma [J]. *Cancer Cell*, 2012, 21(3): 418-29.

[64] KAYAMORI K, KATSUBE K, SAKAMOTO K, et al. NOTCH3 Is Induced in Cancer-Associated Fibroblasts and Promotes Angiogenesis in Oral Squamous Cell Carcinoma [J]. *PLoS One*, 2016, 11(4): e0154112.

[65] WANG Y, JING Y, DING L, et al. Epiregulin reprograms cancer-associated fibroblasts and facilitates oral squamous cell carcinoma invasion via JAK2-STAT3 pathway [J]. *J Exp Clin Cancer Res*, 2019, 38(1): 274.

[66] CHIA-HUEI LEE S-H S, KO-JIUNN LIU, PEI-YI CHU, WEN-CHAN YANG, PINPIN LIN, WAN-YU SHIEH. Interleukin-1 beta transactivates epidermal growth factor receptor via the CXCL1-CXCR2 axis in oral cancer [J]. *Oncotarget*, 2015, 6(36): 38866-80.

[67] GUIMARAES V S N, VIDAL M T A, DE FARO VALVERDE L, et al. Hedgehog pathway activation in oral squamous cell carcinoma: cancer-associated fibroblasts

exhibit nuclear GLI-1 localization [J]. *J Mol Histol*, 2020, 51(6): 675-84.

[68] YANG J, SHI X, YANG M, et al. Glycolysis reprogramming in cancer-associated fibroblasts promotes the growth of oral cancer through the lncRNA H19/miR-675-5p/PFKFB3 signaling pathway [J]. *Int J Oral Sci*, 2021, 13(1): 12.

[69] COSTEA D E, HILLS A, OSMAN A H, et al. Identification of two distinct carcinoma-associated fibroblast subtypes with differential tumor-promoting abilities in oral squamous cell carcinoma [J]. *Cancer Res*, 2013, 73(13): 3888-901.

[70] HASSONA Y, CIRILLO N, HEESOM K, et al. Senescent cancer-associated fibroblasts secrete active MMP-2 that promotes keratinocyte dis-cohesion and invasion [J]. *Br J Cancer*, 2014, 111(6): 1230-7.

[71] CUSTODIO M, BIDDLE A, TAVASSOLI M. Portrait of a CAF: The story of cancer-associated fibroblasts in head and neck cancer [J]. *Oral Oncol*, 2020, 110(104972).

[72] STEINBICHLER T B, SAVIC D, DEJACO D, et al. Pleiotropic Effects of Epithelial Mesenchymal Crosstalk on Head and Neck Cancer: EMT and beyond [J]. *Cancer Microenviron*, 2019, 12(2-3): 67-76.

[73] LINDA ZIANI, SALEM CHOUAIB, THIERY J. Alteration of the Antitumor Immune Response by Cancer-Associated Fibroblasts [J]. *Front Immunol*, 2018, 9(414).

[74] YU B, WU K, WANG X, et al. Periostin secreted by cancer-associated fibroblasts promotes cancer stemness in head and neck cancer by activating protein tyrosine kinase 7 [J]. *Cell Death Dis*, 2018, 9(11): 1082.

[75] CARMELIET P, JAIN R K. Molecular mechanisms and clinical applications of angiogenesis [J]. *Nature*, 2011, 473(7347): 298-307.

[76] WANG S, MA N, KAWANISHI S, et al. Relationships of alpha-SMA-positive fibroblasts and SDF-1-positive tumor cells with neoangiogenesis in nasopharyngeal carcinoma [J]. *Biomed Res Int*, 2014, 2014(507353).

- [77]KALLURI R, ZEISBERG M. Fibroblasts in cancer [J]. Nat Rev Cancer, 2006, 6(5): 392-401.
- [78]WANG F T, SUN W, ZHANG J T, et al. Cancer-associated fibroblast regulation of tumor neo-angiogenesis as a therapeutic target in cancer [J]. Oncol Lett, 2019, 17(3): 3055-65.
- [79]INOUE K I, KISHIMOTO S, AKIMOTO K, et al. Cancer-associated fibroblasts show heterogeneous gene expression and induce vascular endothelial growth factor A (VEGFA) in response to environmental stimuli [J]. Ann Gastroenterol Surg, 2019, 3(4): 416-25.
- [80]SAN MARTIN R, BARRON D A, TUXHORN J A, et al. Recruitment of CD34(+) fibroblasts in tumor-associated reactive stroma: the reactive microvasculature hypothesis [J]. Am J Pathol, 2014, 184(6): 1860-70.
- [81]HERRERA A, HERRERA M, GUERRA-PEREZ N, et al. Endothelial cell activation on 3D-matrices derived from PDGF-BB-stimulated fibroblasts is mediated by Snail1 [J]. Oncogenesis, 2018, 7(9): 76.
- [82]UNTERLEUTHNER D, NEUHOLD P, SCHWARZ K, et al. Cancer-associated fibroblast-derived WNT2 increases tumor angiogenesis in colon cancer [J]. Angiogenesis, 2020, 23(2): 159-77.
- [83]DE PALMA M, BIZIATO D, PETROVA T V. Microenvironmental regulation of tumour angiogenesis [J]. Nat Rev Cancer, 2017, 17(8): 457-74.
- [84]CHEN T W, YANG Z G, WANG Q L, et al. Whole tumour quantitative measurement of first-pass perfusion of oesophageal squamous cell carcinoma using 64-row multidetector computed tomography: correlation with microvessel density [J]. Eur J Radiol, 2011, 79(2): 218-23.
- [85]HUANG C, WANG X L, QI F F, et al. Berberine inhibits epithelial-mesenchymal transition and promotes apoptosis of tumour-associated fibroblast-induced colonic

epithelial cells through regulation of TGF-beta signalling [J]. J Cell Commun Signal, 2020, 14(1): 53-66.

[86] WANG S, SU X, XU M, et al. Exosomes secreted by mesenchymal stromal/stem cell-derived adipocytes promote breast cancer cell growth via activation of Hippo signaling pathway [J]. Stem Cell Res Ther, 2019, 10(1): 117.

[87] AO M, FRANCO O E, PARK D, et al. Cross-talk between paracrine-acting cytokine and chemokine pathways promotes malignancy in benign human prostatic epithelium [J]. Cancer Res, 2007, 67(9): 4244-53.

[88] TODARO M, GAGGIANESI M, CATALANO V, et al. CD44v6 is a marker of constitutive and reprogrammed cancer stem cells driving colon cancer metastasis [J]. Cell Stem Cell, 2014, 14(3): 342-56.

[89] HAN D, WANG M, YU Z, et al. FGF5 promotes osteosarcoma cells proliferation via activating MAPK signaling pathway [J]. Cancer Manag Res, 2019, 11(6457-66.

[90] HONGBO LI Q Z, QI WU, YAYUN CUI, HONG ZHU, MINGMING FANG, XIFA ZHOU, ZHIQIANG SUN, JINGPING YU. Interleukin-22 secreted by cancer-associated fibroblasts regulates the proliferation and metastasis of lung cancer cells via the PI3K-Akt-mTOR signaling pathway [J]. Am J Transl Res, 2019, 11(7): 4077-88.

[91] MA H, WANG J, ZHAO X, et al. Periostin Promotes Colorectal Tumorigenesis through Integrin-FAK-Src Pathway-Mediated YAP/TAZ Activation [J]. Cell Rep, 2020, 30(3): 793-806 e6.

[92] ZHANG M, SHI R, GUO Z, et al. Cancer-associated fibroblasts promote cell growth by activating ERK5/PD-L1 signaling axis in colorectal cancer [J]. Pathol Res Pract, 2020, 216(4): 152884.

[93] KADERA B E, LI L, TOSTE P A, et al. MicroRNA-21 in pancreatic ductal adenocarcinoma tumor-associated fibroblasts promotes metastasis [J]. PLoS One, 2013, 8(8): e71978.

- [94]TAO WANG F N, ROYA NAVAB, JOELLA JOSEPH , EMIN IBRAHIMOV , JING XU , CHANG-QI ZHU,AYELET BORGIDA , STEVEN GALLINGER 6, MING-SOUND TSAO. Senescent Carcinoma-Associated Fibroblasts Upregulate IL8 to Enhance Prometastatic Phenotypes [J]. Mol Cancer Res, 2017, 15(1): 3-14.
- [95]NORTON J, FOSTER D, CHINTA M, et al. Pancreatic Cancer Associated Fibroblasts (CAF): Under-Explored Target for Pancreatic Cancer Treatment [J]. Cancers (Basel), 2020, 12(5):
- [96]ELMUSRATIA A, PILBOROUGH A E, KHURRAM S A, et al. Cancer-associated fibroblasts promote bone invasion in oral squamous cell carcinoma [J]. Br J Cancer, 2017, 117(6): 867-75.
- [97]SATO K, LEE J W, SAKAMOTO K, et al. RANKL synthesized by both stromal cells and cancer cells plays a crucial role in osteoclastic bone resorption induced by oral cancer [J]. Am J Pathol, 2013, 182(5): 1890-9.
- [98]TAKAHASHI H, SAKAKURA K, KAWABATA-IWAKAWA R, et al. Immunosuppressive activity of cancer-associated fibroblasts in head and neck squamous cell carcinoma [J]. Cancer Immunol Immunother, 2015, 64(11): 1407-17.
- [99]HUANG Y H, CHANG C Y, KUO Y Z, et al. Cancer-associated fibroblast-derived interleukin-1beta activates protumor C-C motif chemokine ligand 22 signaling in head and neck cancer [J]. Cancer Sci, 2019, 110(9): 2783-93.
- [100] CHO H, SEO Y, LOKE K M, et al. Cancer-Stimulated CAFs Enhance Monocyte Differentiation and Protumoral TAM Activation via IL6 and GM-CSF Secretion [J]. Clin Cancer Res, 2018, 24(21): 5407-21.
- [101] AL-ANSARI M M, HENDRAYANI S F, SHEHATA A I, et al. p16(INK4A) represses the paracrine tumor-promoting effects of breast stromal fibroblasts [J]. Oncogene, 2013, 32(18): 2356-64.
- [102] FLAVELL R A, SANJABI S, WRZESINSKI S H, et al. The polarization of

immune cells in the tumour environment by TGFbeta [J]. *Nat Rev Immunol*, 2010, 10(8): 554-67.

[103] FEI XING J S, KOUNOSUKE WATABE. Cancer associated fibroblasts (CAFs) in tumor microenvironment [J]. *Front Biosci (Landmark Ed)*, 2010, 15(166-79.

[104] DVORAK H F. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing [J]. *N Engl J Med*, 1986, 315(26): 1650-9.

[105] YU Y, KE L, LV X, et al. The prognostic significance of carcinoma-associated fibroblasts and tumor-associated macrophages in nasopharyngeal carcinoma [J]. *Cancer Manag Res*, 2018, 10(1935-46.

[106] KAMOLPORN AMORNSUPAK T I, PETI THUWAJIT, PORNCHEI O-CHAROENRAT, SUZANNE A ECCLES, CHANITRA THUWAJIT. Cancer-associated fibroblasts induce high mobility group box 1 and contribute to resistance to doxorubicin in breast cancer cells [J]. *BMC Cancer*, 2014, 14(955.

[107] LOUAULT K, BONNEAUD T L, SEVENO C, et al. Interactions between cancer-associated fibroblasts and tumor cells promote MCL-1 dependency in estrogen receptor-positive breast cancers [J]. *Oncogene*, 2019, 38(17): 3261-73.

[108] ZHANG L, YAO J, LI W, et al. Micro-RNA-21 Regulates Cancer-Associated Fibroblast-Mediated Drug Resistance in Pancreatic Cancer [J]. *Oncol Res*, 2018, 26(6): 827-35.

[109] LOTTI F, JARRAR A M, PAI R K, et al. Chemotherapy activates cancer-associated fibroblasts to maintain colorectal cancer-initiating cells by IL-17A [J]. *J Exp Med*, 2013, 210(13): 2851-72.

[110] KADEL D, ZHANG Y, SUN H R, et al. Current perspectives of cancer-associated fibroblast in therapeutic resistance: potential mechanism and future strategy [J]. *Cell Biol Toxicol*, 2019, 35(5): 407-21.

[111] MUTGAN A C, BESIKCIOGLU H E, WANG S, et al. Insulin/IGF-driven

cancer cell-stroma crosstalk as a novel therapeutic target in pancreatic cancer [J]. *Mol Cancer*, 2018, 17(1): 66.

[112] WU X, RUAN L, YANG Y, et al. Analysis of gene expression changes associated with human carcinoma-associated fibroblasts in non-small cell lung carcinoma [J]. *Biol Res*, 2017, 50(1): 6.

[113] QIN X, GUO H, WANG X, et al. Exosomal miR-196a derived from cancer-associated fibroblasts confers cisplatin resistance in head and neck cancer through targeting CDKN1B and ING5 [J]. *Genome Biol*, 2019, 20(1): 12.

[114] AFFOLTER A, SCHMIDTMANN I, MANN W J, et al. Cancer-associated fibroblasts do not respond to combined irradiation and kinase inhibitor treatment [J]. *Oncol Rep*, 2013, 29(2): 785-90.

[115] TASNUVA D KABIR R J L, HATAITIP TASENA , MASSIMILIANO MELLONE , RICARDO D COLETTA , ERIC K PARKINSON , STEPHEN S PRIME , GARETH J THOMAS , IAN C PATERSON , DONGHUI ZHOU , JOHN MCCALL , PAUL M SPEIGHT , DANIEL W LAMBERT. A miR-335/COX-2/PTEN axis regulates the secretory phenotype of senescent cancer-associated fibroblasts [J]. *Aging (Albany NY)*, 2016, 8(8): 1608-35.

[116] MASSIMILIANO MELLONE C J H, STEVE THIRDBOROUGH , TOBY MELLOWS , EDWIN GARCIA , JEONGMIN WOO , JOANNE TOD , STEVE FRAMPTON , VERONIKA JENEI , KARWAN A MOUTASIM , TASNUVA D KABIR , PETER A BRENNAN , GIULIA VENTURI , KIRSTY FORD , NICOLAS HERRANZ , KUE PENG LIM , JAMES CLARKE , DANIEL W LAMBERT, STEPHEN S PRIME , TIMOTHY J UNDERWOOD , PANDURANGAN VIJAYANAND , KEVIN W ELICEIRI , CHRISTOPHER WOELK , EMMA V KING , JESUS GIL , CHRISTIAN H OTTENSMEIER , GARETH J THOMAS Induction of fibroblast senescence generates a non-fibrogenic myofibroblast phenotype that

differentially impacts on cancer prognosis [J]. Aging (Albany NY), 2016, 9(1): 114-32.

[117] EMMA L JAMES R D M, GAYANI N PITIYAGE, ALICE M DE CASTRO, KATIE S VIGNOLA , JANICE JONES , ROBERT P MOHNEY , EDWARD D KAROLY , STEPHEN S PRIME, ERIC KENNETH PARKINSON. Senescent human fibroblasts show increased glycolysis and redox homeostasis with extracellular metabolomes that overlap with those of irreparable DNA damage, aging, and disease [J]. J Proteome Res, 2015, 14(4): 1854-71.

[118] FANG J, XIAO L, JOO K I, et al. A potent immunotoxin targeting fibroblast activation protein for treatment of breast cancer in mice [J]. Int J Cancer, 2016, 138(4): 1013-23.

[119] OHSHIO Y, TERAMOTO K, HANAOKA J, et al. Cancer-associated fibroblast-targeted strategy enhances antitumor immune responses in dendritic cell-based vaccine [J]. Cancer Sci, 2015, 106(2): 134-42.

[120] AL-JOMAH N, AL-MOHANNA F H, ABOUSSEKHRA A. Tocilizumab suppresses the pro-carcinogenic effects of breast cancer-associated fibroblasts through inhibition of the STAT3/AUF1 pathway [J]. Carcinogenesis, 2021, 42(12): 1439-48.

[121] SIEFKER-RADTKE A O, LORIOT Y. Erdafitinib for locally advanced or metastatic urothelial carcinoma [J]. Am J Health Syst Pharm, 2022, 79(11): 824-5.

[122] KIM R D, SARKER D, MEYER T, et al. First-in-Human Phase I Study of Fisogatinib (BLU-554) Validates Aberrant FGF19 Signaling as a Driver Event in Hepatocellular Carcinoma [J]. Cancer Discov, 2019, 9(12): 1696-707.

[123] MARGIT A HUBER N K, JOHN E PARK, ROLAND D SCHUBERT, WOLFGANG J RETTIG, RALF U PETER, PILAR GARIN-CHESA. Fibroblast activation protein: differential expression and serine protease activity in reactive stromal fibroblasts of melanocytic skin tumors [J]. J Invest Dermatol, 2003, 120(2): 182-8.

- [124] OSTERMANN E, GARIN-CHESA P, HEIDER K H, et al. Effective immunoconjugate therapy in cancer models targeting a serine protease of tumor fibroblasts [J]. Clin Cancer Res, 2008, 14(14): 4584-92.
- [125] S WELT, C R DIVGI, A M SCOTT, et al. Antibody targeting in metastatic colon cancer: a phase I study of monoclonal antibody F19 against a cell-surface protein of reactive tumor stromal fibroblasts [J]. J Clin Oncol, 1994, 12(6): 1193-203.
- [126] OZDEMIR B C, PENTCHEVA-HOANG T, CARSTENS J L, et al. Depletion of carcinoma-associated fibroblasts and fibrosis induces immunosuppression and accelerates pancreas cancer with reduced survival [J]. Cancer Cell, 2014, 25(6): 719-34.
- [127] ZALCMAN G, MAZIERES J, MARGERY J, et al. Bevacizumab for newly diagnosed pleural mesothelioma in the Mesothelioma Avastin Cisplatin Pemetrexed Study (MAPS): a randomised, controlled, open-label, phase 3 trial [J]. The Lancet, 2016, 387(10026): 1405-14.